IN THE SPOTLIGHT

The gRASs Is Greener: Potential New Therapies in Lung Cancer with Acquired Resistance to KRAS^{G12C} Inhibitors

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Summary: Inhibitors of KRAS^{G12C} that bind the target in its inactive conformation and lock it in off-mode have shown early signs of clinical activity in patients with *KRAS*^{G12C}-mutant lung cancer, but responses tend to be short-lived and invariably prelude the development of acquired resistance through largely unexplored mechanisms. A new study describes the emergence of RAS-MAPK heterogeneous subclonal alterations in a patient relapsed on a KRAS^{G12C} inactive-state inhibitor and identifies a novel KRAS^{Y96D}-resistant variant that is druggable by a next-generation compound capable of associating with KRAS^{G12C} in its active configuration.

See related article by Tanaka et al., p. 1913 (8).

KRAS and the structurally related NRAS and HRAS GTPases relay mitogenic stimuli from the extracellular environment to the cell nucleus by stimulating a series of cytoplasmic kinases (RAF, MEK, and ERK, collectively defining the MAPK cascade) that culminates in the stabilization and activation of transcription factors driving cell-cycle progression (1). For reversible implementation of this pathway, RAS proteins oscillate between an active, GTP-bound and an inactive, GDP-bound state at rates controlled by upstream growth factor-dependent signals. RAS family oncogenic mutations are common in tumors and typically result in single amino acid substitutions that constitutively activate the encoded enzymes by thwarting their ability to hydrolyze GTP, hence compromising catalytic autoinhibition. KRAS mutations, in particular, are found in lung, pancreatic, and colorectal cancers at frequencies of about 30%, 90%, and 40%, respectively (2).

Effective targeting of KRAS has proved daunting due to its high affinity for GTP and lack of sufficiently large pockets that enable accommodation of allosteric inhibitors. Moreover, pharmacologic interception of KRAS downstream effectors—namely, the MAPK cascade—is usually counteracted by feedback signal compensation (3). These limitations notwithstanding, the promise of KRAS inactivation has been recently revived by the discovery of inhibitors that selectively target KRAS proteins harboring a glycine-to-cysteine mutation at position 12 (G12C). Such small molecules covalently bind the mutated cysteine and occupy a pocket in the so-called switch-II region when KRAS^{G12C} is in its inactive, GDPbound conformation, thus abrogating RAS-dependent sig-

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naling. Inactive-state KRAS^{G12C} inhibitors can do so because the mutant protein, although mostly engaged in its active conformation, still undergoes nucleotide cycling and experiences periods of inactivity, which allows for drug trapping and covalent attack (4).

Findings from recently completed and ongoing phase I/II trials are a testimony to the merits-but also a warning of the shortcomings-of targeting KRASG12C. When tested in patients with KRASG12C-mutant metastatic non-small cell lung cancer (NSCLC), the $\ensuremath{\mathsf{KRAS}^{\text{G12C}}}$ inhibitor sotorasib (AMG 510) was efficacious, with an overall response rate of 32.2% and a median progression-free survival of 6.3 months (5). Likewise, the objective response rate of patients with advanced or metastatic NSCLC treated with adagrasib (MRTX849, another inactive-state KRASG12C inhibitor characterized by a long half-life that equals the 24-hour synthesis rate of the KRAS protein) was 45% (6). Although disease control was remarkable in both studies, a relatively large fraction of patients responded suboptimally to either therapy, and many of those who had received some benefit relapsed quickly. According to preclinical experiments in isogenic cell lines, poor response *ab initio* (known as primary resistance) might be explained with a rapid process of nonuniform adaptation whereby some cells escape inhibition by producing new KRASG12C (which is promptly converted to the active, drug-refractory state) while others without sufficient expression of newly synthetized KRASG12C are eliminated by treatment (7). Less is known about the mechanisms underlying acquired resistance, and whether they mainly involve selection of genetically resistant subclones or plastic fitness variations.

In this issue of *Cancer Discovery*, Tanaka and colleagues (8) begin to delineate genetic alterations that may be responsible for the acquisition of secondary resistance in the clinic and illustrate potential therapeutic opportunities to target some of them. They describe a patient with metastatic NSCLC positive for the *KRAS*^{G12C} mutation who was treated with adagrasib. The patient had an initial objective response (32% reduction in tumor size) but showed evidence of progressive disease after approximately 4 months of treatment.

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Figure 1. The increasingly populated landscape of potential therapeutic options (butterflies, in red) for KRAS^{G12C}-mutant NSCLC with acquired resist-ance to inactive-state KRAS^{G12C} inhibitors, tailored around the genomic characteristics of resistance mutations (grass and tree leaves, in black; undruggable mutations are in leafless branches). "Insects and Grasses" by Yamamoto Baiitsu (1783–1856), adapted and modified from Creative Commons under a Creative Commons Attribution CC0 1.0 (Universal Public Domain Dedication).

Comparative analysis of cell-free DNA (cfDNA) before treatment and at the onset of resistance revealed the persistence of the KRASG12C mutation and the appearance of many distinct new mutations, all giving rise to protein products that are not druggable by inactive-state KRAS^{G12C} inhibitors. These alterations are predicted to converge on the reactivation of the RAS-MAPK pathway and include gain-of-function mutations in KRAS (which likely originated in trans in the remaining wild-type gene copy), NRAS, BRAF, and MAP2K1 (encoding the MEK1 protein; Fig. 1).

An interesting piece of information is the discovery of a novel, previously unidentified tyrosine-to-aspartate mutation at position 96 of KRAS (KRAS^{Y96D}; Fig. 1). On the basis of the crystal structure of different inactive-state KRASG12C inhibitors bound to KRAS^{G12C}, the Y96D substitution appears to disrupt a critical hydrogen bond between the hydroxyl group of tyrosine 96 and the pyrimidine ring of adagrasib. More generally, the amino acid change at the tyrosine 96 locus is thought to weaken drug-target chemical interactions by making the switch-II pocket of the mutant enzyme more hydrophilic. This modification also affects target occupancy by KRAS^{G12C} inhibitors other than adagrasib, thus representing a shared liability of currently available compounds. Consistent with a functional role of KRAS^{Y96D}, ectopic introduction of the mutant gene into KRASG12C-addicted cancer cell lines attenuated the growth-suppressing effect of inactive-state KRAS^{G12C} inhibitors and enhanced RAS signaling, indicating that KRAS^{Y96D} is an oncogenic mutation that leads to constitutive RAS activation and imparts resistance to KRASG12C blockade.

Of note, the allele frequency of the KRASG12C mutation in the posttreatment cfDNA was much higher than that of the newly emerging alterations, pointing to KRASG12C as a truncal mutation that is not extinguished by treatment and dominates over minor subclonal branches harboring the putative resistance alterations. Given the very low prevalence (also cumulatively) of the identified mutations, their causal role in establishing tumor progression and clinical relapse is not immediately evident. However, cfDNA values hardly allow for inferring the relative contribution of the different subclonal mutations to the genomic architecture of the tumor, and it may well be that the representation of mutant DNA was more prominent-therefore, more pervasive in dictating resistance-in the lesions carried by the patient than in blood. It may also be that paracrine growth factors secreted by the tiny portion of resistant cells protected the surrounding arrays of sensitive cells from the therapeutic insult. Finally, in the absence of ultradeep multiregion sequencing data on the pretreatment tumor tissues, it remains unclear whether the mutant subpopulations preexisted at very low frequency in the original tumor or materialized de novo during treatment.

Can one envisage therapeutic options to overcome acquired resistance to inactive-state KRAS^{G12C} inhibitors? Importantly, Tanaka and colleagues (8) show that a new compound targeting active, GTP-bound KRASG12C retains potency against KRAS^{Y96D} (Fig. 1). This drug, called RM-018, has affinity for the chaperone protein cyclophilin-A. The resulting complex facilitates the formation of extensive protein-protein surface interactions that sterically occlude KRASG12C in its active state and preclude KRAS association with downstream signaling effectors. When tested in KRASG12C-mutant cell lines with exogenous expression of KRAS^{Y96D}, RM-108 markedly impaired cell proliferation and reduced RAS signaling. This is welcome evidence that at least one mechanism of therapeutic resistance could be tamed pharmacologically, although it will be crucial to extend these initial observations from engineered cells to *in vitro* and *in vivo* models in which *KRAS*^{G12C} and *KRAS*^{Y96D} mutations spontaneously arise during the tumor natural history.

Tracking down an individual therapy covering the plethora of heterogeneous mutant proteins documented in the study by Tanaka and colleagues (8) will likely be problematic, especially considering that the identified mutations in KRAS and NRAS (with the exception of KRAS^{Y96D}) are not actionable. Nonetheless, some of the reported mutations (specifically, those detected in the BRAF and MAP2K1 genes) result in proteins that are vulnerable to pharmacologic neutralization, which bodes well for dual therapies against inactive or active KRAS^{G12C} together with BRAF or MEK inhibitors (Fig. 1). Fortunately enough, inactive-state KRASG12C drugs are well tolerated (5, 6), with no dose-limiting toxicities or grade 4 therapy-related adverse events. Therefore, a further opportunity could be the design of multiple combination therapies in which a common anti-KRAS^{G12C} backbone is combined with treatments that affect the MAPK cascade more profoundly than single-agent BRAF or MEK blockade, for example through vertical inactivation of both BRAF and MEK or by including ERK inhibitors (Fig. 1). At least in principle, concomitant shrinkage of the dominant bulk of KRASG12C-mutant cells together with the MAP2K1- and BRAF-mutant minor subclones might engender a "cascade effect" on the growth dynamics of other mutant subclones present in the tumor ecosystem, potentially leading to extinction or at least contraction of drug-resistant foci fueled by currently undruggable non-G12C KRAS or NRAS mutations (9). Clonal variations in the genetic composition of treated tumors may also modify the synthetic rate of newly produced KRASG12C and the ratio between active and inactive RAS in functionally heterogeneous tumor subpopulations, which may influence adaptive fitness and susceptibility to inactive-state inhibitors. As done with other targeted therapies in different tumor contexts, preemptive strategies aimed at using inhibitors against the resistance oncoproteins as up-front therapies, before clinical manifestation of the corresponding mutations, should be considered as well (10).

More work is needed to better understand the population prevalence, biological relevance, and therapeutic exploitability of the proposed resistance mechanisms, and it is difficult to anticipate whether laboratory results will be successfully translated into clinical benefit for patients with *KRAS*^{G12C}mutant cancer. At the same time, much work has also already been done. Until only a couple of years ago, the land of opportunities for effective and durable treatment of *KRAS*-mutant tumors was inaccessible and desolate. With a growing body of knowledge on the genetic determinants of acquired resistance to inactive-state KRAS^{G12C} inhibitors and an expanding arsenal of different classes of KRAS^{G12C}-targeting agents, this land is more fecund now, and yields blades of greener grass.

Authors' Disclosures

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REFERENCES

- Pylayeva-Gupta Y, Grabocka E, Bar-Sagi D. RAS oncogenes: weaving a tumorigenic web. Nat Rev Cancer 2011;11:761–74.
- 2. Simanshu DK, Nissley DV, McCormick F. RAS proteins and their regulators in human disease. Cell 2017;170:17-33.
- Trusolino L, Bertotti A. Compensatory pathways in oncogenic kinase signaling and resistance to targeted therapies: six degrees of separation. Cancer Discov 2012;2:876–80.
- Moore AR, Rosenberg SC, McCormick F, Malek S. RAS-targeted therapies: is the undruggable drugged? Nat Rev Drug Discov 2020;19: 533–52.
- Hong DS, Fakih MG, Strickler JH, Desai J, Durm GA, Shapiro GI, et al. KRAS(G12C) inhibition with sotorasib in advanced solid tumors. N Engl J Med 2020;383:1207–17.
- 6. Poh A. Another KRAS inhibitor holds its own. Cancer Discov 2020;10:OF2.
- Xue JY, Zhao Y, Aronowitz J, Mai TT, Vides A, Qeriqi B, et al. Rapid non-uniform adaptation to conformation-specific KRAS(G12C) inhibition. Nature 2020;577:421–5.
- Tanaka N, Lin JJ, Li C, Ryan MB, Zhang J, Kiedrowski LA, et al. Clinical acquired resistance to KRAS^{G12C} inhibition through a novel KRAS switch-II pocket mutation and polyclonal alterations converging on RAS-MAPK reactivation. Cancer Discov 2021;11:1913–22.
- 9. Pogrebniak KL, Curtis C. Harnessing tumor evolution to circumvent resistance. Trends Genet 2018;34:639–51.
- Bivona TG, Doebele RC. A framework for understanding and targeting residual disease in oncogene-driven solid cancers. Nat Med 2016;22:472-8.

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